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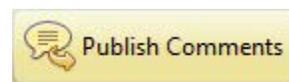


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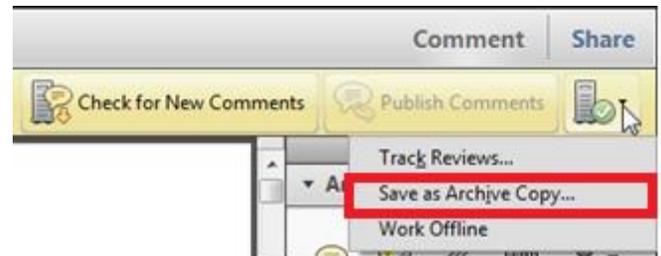
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Effects of the combination of gamma irradiation and Ag nanoparticles polyethylene films on the quality of fresh bottom mushroom (*Agaricus bisporus* L.)

Mahdi Ghasemi-Varnamkhashti | Ayat Mohammad-Razdari | Seyedeh Hoda Yoosefian | Zahra Izadi

Department of Mechanical Engineering of Biosystems, Shahrekord University, Shahrekord, Iran

Correspondence

Mahdi Ghasemi-Varnamkhashti, Department of Mechanical Engineering of Biosystems, Shahrekord University, Shahrekord, Iran. Email: ghasemymahdi@ut.ac.ir; ghasemy-mahdi@gmail.com

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Abstract

This study was conducted to examine the combined method of gamma irradiation doses (0, 1, and 2 kGy) and Ag nanoparticles polyethylene films on the quality of fresh bottom mushroom during storage. For this purpose, physical and chemical properties such as pH, color, weight loss, as well as texture parameters test of the mushroom samples were measured and microbial test for Ag nanoparticles polyethylene films were also performed during 21 days of storage at 4°C. It was observed that the samples irradiated with a dose of 2 kGy and placed in Ag nanoparticles polyethylene films had the lowest reduction in pH (14.33%) and L* (lightness; 6.0%), while weight loss, b* and browning index had the fewest changes with the amount of 9.47, 5.58, and 13.84, respectively. Also, a*, for the control sample and Ag nanoparticles polyethylene films after 21 days of storage increased up to 8.39 and 7.17%, respectively, compared to the initial samples. Also, the greatest changes in the firmness and elasticity for the treatment, respectively was 5.22 and 3.24% compared to the initial samples. Finally, it has been indicated that Ag nanoparticles polyethylene films could prevent the accumulation of microbial load. The results thus demonstrate that the combined use of gamma irradiation and Ag nanoparticles polyethylene films is an effective approach to maintain the quality of fresh bottom mushroom during storage.

Practical applications

Irradiating food causes changes in flavor, color, nutrients, taste, and other qualitative properties and such merits could extend the shelf life of the food products for preservation aims. Also, use of nanoparticles polyethylene films could help to better preservation of the mushrooms. Such combinations (nanoparticles films with gamma rays) could be of interest for the industry in packaging process and consequently export for long time consumption.

1 | INTRODUCTION

Bottom mushroom (*Agaricus bisporus*) is one of the most commonly used type of mushroom worldwide and makes about 40% of the world mushroom production (Guan, Fan, & Yan, 2013). Storage duration of fresh mushrooms is very short and is customer-friendly until it does

not change the quality and freshness (Oliveira, Sousa-Gallagher, Mahajan, & Teixeira, 2012a). Mushrooms quality attributes include browning, softening (Yurttas, Moreira, & Castell-Perez, 2014), cap widening, and losses in weight (Kim, Ko, Lee, Park, & Hanna, 2006).

Many ways to keep the freshness and quality of fresh mushrooms during storage have been reported, such as electron irradiation (Mami, Peyvast, Ziaie, Ghasemnezhad, & Salmanpour, 2014), packaging with different films (Taghizadeh, Gowen, Ward, & O'Donnell, 2010), packaging with cinnamon oil (Echegoyen & Nerin, 2015), packaging with modified atmosphere (Kim et al., 2006), washed with hydroxide peroxide (Sapers, Miller, Choi, & Cooke, 1999), and ozone (Yuk, Yoo, Yoon,

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Marshall, & Oh, 2007). These ways are very efficient to protect mushroom texture and quality of this product (Gilman, Jacksens, De-Meulenaer, & Devlieghere, 2015).

Irradiating food causes changes in flavor, color, nutrients, taste, and other qualitative properties (Oliveira et al., 2012a). The use of gamma irradiation has long history in different types of food such as citrus (Mahrouz et al., 2002; Oufedjikh, Mahrouz, Amiot, & Lacroix, 2000; Oufedjikh, Mahrouz, Lacroix, Amiot, & Taccini, 1998), Spices (Khatun et al., 2017), and vegetables (Majeed et al., 2017). Also gamma irradiation because of high penetration power is commonly used in food packaging that of course it is considered as a method for cold sterilization (Madera-Santana, Meléndrez, González-García, Quintana-Owen, & Pillai, 2016). The level of changes caused by the ray in different foods in the aroma, color, and taste depends on the food material, irradiation dose, and ray source (Oliveira, Sousa-Gallagher, Mahajan, & Teixeira, 2012b).

In a study conducted by Han et al. (2015) for increasing shelf-life of the wild mushroom, poly (lactic acid [PLA]) packaging films were used and the results demonstrated that the shelf-life has increased up to 18 days and this kind of film showed antimicrobial activity. Furthermore, Qin et al. (2015) conducted a study on the effect of antibacterial film of PLA/poly (ϵ -caprolactone) (PCL) on the physicochemical and microbial properties of bottom mushroom. The results showed that packaging bottom mushroom with PLA polymer films maintained the color and sensory and physical properties and prevented water vapor loss from the mushroom after 12 days of storage and microbial load aggregation. Gantner et al. (in press) conducted a study on the effect of type of packaging films and modified atmosphere on the shelf-life of white mushroom. According to the results, after 14-day storage, a polymer film in combination with modified atmosphere maintained the color, weight loss, texture, and shelf-life. Donglu et al. (2016) conducted a study on the effect of polyethylene (PE) film on mushroom shelf-life and concluded that this type of film maintained the mushroom shelf-life and quality and played a significant role in commercialization of the product.

Mushroom is one of the most popular foods, but its customer satisfaction is for healthy and white, nonshrink, and nonbrown warheads. In contrast, this product is highly corrupted and its qualitative changes decrease in a short time. This product is dramatically produced around the world (Xu et al., 2017). Irradiation is a process confirmed by global health organizations and exist irradiation companies for agricultural and food products in all countries and irradiation of a high volume of the product is very cost effective (Ekezie, Cheng, & Sun, 2018). Furthermore, the polyethylene coating is a polymer coating of silver nanoparticles that have very low cost in high production volume. The silver nanoparticles contained in the coatings in combination with irritation of products eliminate the use of chemicals and have little disadvantage over other methods of storage. The cost of deterioration of high volume of the product as well as the cost of treatment caused by the introduction of various chemicals into the human body is far more than packaging with this type of coating and is very cost effective in a large volume.

Some studies have been conducted on the mushroom by just gamma irradiation method (Charlesby, 2016; Choi, Park, Choi, Kim, & Chun, 2015; Marra et al., 2016; Schmid, Held, Hammann, Schlemmer, & Noller, 2015; Severino et al., 2015). But, regarding the knowledge of the paper authors, so far no study has been reported on the combination of gamma irradiation and Ag nanoparticles polyethylene films. The aim of this study is to evaluate the effect of different doses of gamma irradiation in combination with Ag nanoparticles polyethylene films on the physical and chemical properties and texture of fresh bottom mushroom and microbial properties of Ag nanoparticles polyethylene films. Therefore, the idea behind of the research is quit novel and original.

2 | MATERIALS AND METHODS

2.1 | Samples preparation and irradiation

The samples of fresh mushrooms were harvested from the farms in 2016 with uniform size, same color, and no injuries. Specifications of gamma source were gamma cell (GC) 220, Nordin, dose rate 3.05 Gy, 18 kKori Source power, 0 (control), 1 and 2 kGy irradiation dose (Fernandes et al., 2016). Then, they were stored at 4°C, and the experiments were performed.

2.2 | Fabricating and producing the films

Medium-density polyethylene film (1.2 kg/m³) was prepared. Ag nanoparticles with a size of 35 nm were also purchased. To combine nanoparticles with a polymer film, extrusion process took place in the extruder. The temperature in different areas of extruder, from feeding chamber to output was 125, 145, 155, 170, 185, 195, and 200°C, respectively. The extruder chamber pressure was 12.5 bar and melt temperature was about 200°C.

To ensure the proper path and conditions, polyethylene film and Ag nanoparticles (0.5 and 1 wt %) were well mixed and fed through a funnel into the extruder. The materials were mixed together by creating shear force and pressure. The mixture was exited from the extruder, and the created granules was then exited from the chamber, after being heated it was passed as a thin film over a cooling roller and subsequently threw in a cold water pool (Emamifar, Kadivar, Shahedi, & Soleimani-Zad, 2010).

The treatments of Ag nanoparticles polyethylene films and irradiation were as follow:

- Nonirradiated samples in paper bags ©
- Nonirradiated samples in polyethylene films without Ag nanoparticles (PE + C)
- 1 kGy irradiated sample in polyethylene films without Ag nanoparticles (PE + 1i)
- 2 kGy irradiated sample in polyethylene films without Ag nanoparticles (PE + 2i)
- Nonirradiated samples in Ag nanoparticles polyethylene films (PE + Ag + C)

- 147 • 1 kGy irradiated samples in Ag nanoparticles polyethylene films
- 148 (PE + Ag + 1i)
- 149 • 2 kGy irradiated samples in Ag nanoparticles polyethylene films
- 150 (PE + Ag + 2i)

151 **2.3 | pH measurement**

152 After separating the waste over the mushroom cap, 20 fresh mush-
 153 rooms per treatment were cut into small pieces, mixed well by a
 154 blender and passed through a clean fabric. Finally, pH level of the solu-
 155 tion was measured by a pH meter (PH-2211, Hana, Italy) (Aday, Caner,
 156 & Rahvali, 2011).

157 **2.4 | Measuring color of samples**

158 The color of samples was measured using a portable colorimeter (Kon-
 159 ica Minolta, CR400, Japan). To calibrate the colorimeter the standard
 160 white plate (CR-A43) was used and the parameters L^* (lightness), a^*
 161 (red–green), and b^* (yellow–blue) were recorded. Browning index (BI)
 162 was calculated using the following equations (Abbasi & Azari, 2009):

$$BI = \frac{[100(x-0.31)]}{0.17} \tag{1}$$

$$x = \frac{(a^* + 1.75L^*)}{(5.645L^* + a^* - 3.012b^*)} \tag{2}$$

163 **2.5 | Weight loss**

164 The mushroom weight for each treatment was recorded at the begin-
 165 ning and end of the experiments. Before the experiment, all the treat-
 166 ments were labeled and the sample weight loss percentage was
 167 recorded (Koutsimanis, Harte, & Almenar, 2015).

168 **2.6 | Texture analysis of the samples**

169 Twenty mushrooms were used and Texture Test (TPA) was performed
 170 on mushroom cap using Instron (STM-Santam20, Iran) under the condi-
 171 tions as follow: test speed of 2 mm/s, pretest speed of 10 mm/s, and
 172 30% strain. Then, force-time diagram was calculated using the software
 173 installed on the apparatus and the firmness and elasticity of the mush-
 174 room samples were calculated using the software (Wong et al., 2017).

175 **2.7 | Microbial test**

176 To perform microbial tests, *Escherichia coli* bacteria ATCC 25922 and
 177 *Staphylococcus aureus* ATCC 29523, respectively, were used as
 178 negative-gram and positive-gram bacterial microorganisms. For cultiva-
 179 tion of the microorganisms, Violet Red Bile Dextrose Agar (VRBDA)
 180 medium was used for the bacteria *E. coli* cultivation and Mannitol Salt
 181 Agar (MSA) medium was used for the bacteria *S. aureus* cultivation.
 182 Both sterile nutrients agar were kept until reaching the desired number
 183 for performing microbial tests for 24 hr at 37°C.

184 Polymer film was cut as a small circle with a diameter of 5 cm, dis-
 185 infected with alcohol at 70°C, and 15 ml of the bacteria *E. coli* and *S.*

TABLE 1 Effect of different treatments on pH value during storage

	Storage time (Week)			
	0 Week	1 Week	2 Week	3 Week
C	6.21 ± 0.06	5.64 ± 0.07	5.39 ± 0.01	4.09 ± 0.02
PE + C	6.21 ± 0.06	5.87 ± 0.05	5.60 ± 0.04	4.21 ± 0.05
PE + 1i	6.21 ± 0.06	6.13 ± 0.06	6.13 ± 0.01	6.13 ± 0.04
PE + 2i	6.21 ± 0.06	6.09 ± 0.03	5.82 ± 0.03	4.81 ± 0.06
PE + Ag + C	6.21 ± 0.06	6.01 ± 0.04	5.71 ± 0.07	4.60 ± 0.02
PE + Ag + 1i	6.21 ± 0.06	6.14 ± 0.02	5.97 ± 0.05	5.10 ± 0.03
PE + Ag + 2i	6.21 ± 0.06	6.17 ± 0.05	6.02 ± 0.04	5.32 ± 0.03
Overall	6.21 ± 0.06A	6.10 ± 0.08B	5.73 ± 0.12C	4.86 ± 0.07D

Data are means ± SD of three replicates. A–D means in the same row with different letters are significantly different ($p \leq .05$; mean separation was performed by Tukey test).

aureus was added to the Falcon. The Falcon containing the film and 186
 microorganisms was kept for 24 hr at 37°C. To count the number of 187
 colonies, dilution was done at 10^{-6} and 10^{-7} . Then, using micrometer 188
 sampler, 0.1 ml of microbial suspension was taken and sprayed over 189
 VRBDA for the bacteria *E. coli* cultivation and over MSA for the bacte- 190
 ria *S. aureus* cultivation in the medium, and kept for 24 hr at 37°C. 191
 After 24 hr, the number of colonies was counted and multiplied by the 192
 dilution determined (Restrepo-Florez, Bassi, & Thompson, 2014). 193

194 **2.8 | Data analysis**

In this study, experiments were conducted at three stages and statisti- 195
 cal analysis was performed using software SAS 9.1.3. Additionally, 2- 196
 way ANOVA method was used to examine the effect of different treat- 197
 ments on the mushroom quality and the differences between means 198
 were examined using Tukey model at the significance level of .05. 199

200 **3 | RESULTS AND DISCUSSION**

Table 1 shows the pH value in each treatment during storage that was 201
 6.21 at the beginning of the experiment. pH values for all treatments 202
 significantly reduced by increasing the storage duration. The results are 203
 consistent with the study of Aday (2016) who reported pH value 204
 reduced by increasing the storage duration. pH values influenced by 205
 irradiation dose and Ag nanoparticles were measured during three 206
 weeks of storage. The highest pH value among all treatments belongs 207
 to the sample irradiated in Ag nanoparticles polyethylene films that 208
 O_2/CO_2 value due to reduced respiratory rate compared with the sam- 209
 ples of nonirradiated inside the paper bag and the samples irradiated 210
 with the dose of 1 kGy of polyethylene films without Ag nanoparticles 211
 (PE + 1i) is in the balance. It seems that the production of organic acids 212
 by microorganisms has reduced the pH value in the mushroom 213
 (Oliveira et al., 2012b). 214

Color Index is one of the important parameters for the consumer. 215
 When the mushrooms are harvested white, they began to slowly 216

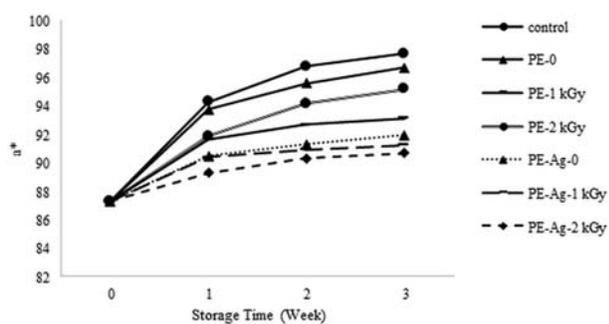


FIGURE 1 Effect of different treatments on a^* value during storage

change color and become dark gradually (Cao et al., 2010). According to the results, during storage, no significant difference was observed between different treatments in the values of a^* , but the treatments irradiated with the dose of 2 kGy in Ag nanoparticles polyethylene films after the storage time were less red and compared to the control treatment in the paper bag were significantly different and less discolored (Figure 1). In all treatments, L^* value reduced and b^* value increased and browning index remained almost stable after two weeks of the storage (Tables 2–4). The study results are consistent with the findings of Caner and Aday (2009) where strawberry samples become dark over time.

Based on the results, a significant difference was observed between the treatments in the packages. Most of changes in the index b^* in the control treatments were observed after the storage period. Conversely, by increasing the irradiation dose, the index b^* value increased and lightness reduced. Yellowing value increased by increasing the irradiation dose because the mushroom lost its volatiles during storage and browning index after the harvest is linearly related to storage time (Anthon & Barrett, 2003). Also, for the samples placed in Ag nanoparticles polyethylene films, Ag nanoparticles prevent the mold growth on the film as well as the color change and increase browning index and b^* . Jo, Son, Shin, and Byun (2003) in their research showed that by increasing the irradiation dose and the placement of the

samples at refrigerator temperature, the value of the index b^* will increase in comparison to the control.

The amount of weight loss in all treatments is shown in Table 5. The mushroom quality reduced over time due to the loss of intracellular water (Khan & Gibbons, 2014). Based on the results, a significant increase was observed for all treatments during storage. The results of statistical analysis showed that no significant difference was found between the samples irradiated with the dose 2 kGy in polyethylene bags with and without Ag nanoparticles and at the end of the storage period, the value of weight loss was 1.5 and .9%, respectively. The highest value of weight loss of 3.02, 1.9, 1.7, and 1.1% were respectively for the control sample, the sample nonirradiated in polyethylene films, the sample irradiated with doses of 1 and 2 kGy in polyethylene film, nonirradiated samples in polyethylene film with Ag nanoparticles. The reason for this phenomenon is that the intracellular water of the mushroom samples reduces by increasing the storage time and becomes dried. Also, due to the respiration of the biological activity, the moisture content reduces (Burton, 1989). Because the cells are in fresh mushrooms, fresh samples have also higher density (Zhou, Lv, He, He, & Shi, 2011).

Texture analysis (TPA) shows important indices for the samples. Texture is the most important parameter that is related to the mechanical and structural properties of food (Abbott & Harker, 2004). During storage, texture parameters, including firmness and elasticity of the mushroom, reduced by increasing adhesion. According to the literature, the firmness is associated with the cell turgor pressure, cell size, cell wall resistance, and intercellular adhesion (Aday, Buyukcan & Caner, 2013).

Figure 2 shows the changes in firmness in the treatments. In general, for the samples irradiated in Ag nanoparticles polyethylene films, the mushroom firmness reduced in comparison with other treatments. The control samples without irradiation in polyethylene bags without Ag nanoparticles had less firmness during storage. Samples of PE-Ag-2 kGy, PE-Ag-1 kGy, PE-2 kGy, and PE-Ag samples have highest firmness at the end of study as 700 ± 23 , 510 ± 31 , 460 ± 39 , and 401 ± 28 , respectively. Reduced turgor pressure on the walls of cells, weight, and

TABLE 2 Effect of different treatments on L^* value during storage

Treatment	Storage time (Week)			
	0 Week	1 Week	2 Week	3 Week
C	73.5 ± 2.29	57.9 ± 2.12	54.1 ± 0.91	45.2 ± 0.78
PE + C	73.5 ± 2.29	69.5 ± 1.19	69.2 ± 1.02	64.4 ± 1.43
PE + 1i	73.5 ± 2.29	70.2 ± 1.27	69.3 ± 1.31	65.3 ± 1.08
PE + 2i	73.5 ± 2.29	71.6 ± 2.02	70.7 ± 1.67	66.5 ± 1.31
PE + Ag + C	73.5 ± 2.29	71.4 ± 0.97	70.4 ± 2.03	65.8 ± 0.82
PE + Ag + 1i	73.5 ± 2.29	72.3 ± 1.37	71.6 ± 1.09	68.3 ± 2.05
PE + Ag + 2i	73.5 ± 2.29	72.7 ± 1.76	72.0 ± 1.15	68.5 ± 2.12
Overall	73.5 ± 2.29 A	70.72 ± 0.83A	68.12 ± 1.52B	57.85 ± 0.93C

Data are means ± SD of three replicates.

A–C means in the same row with different letters are significantly different ($p \leq .05$) (mean separation was performed by Tukey test).

TABLE 3 Effect of different treatments on *b** value during storage

Treatment	Storage time (Week)			
	0 Week	1 Week	2 Week	3 Week
C	34.67 ± 1.52	42.59 ± 1.05	57.63 ± 0.08	76.80 ± 0.07
PE + C	34.67 ± 1.52	39.13 ± 1.12	42.94 ± 0.15	55.19 ± 0.09
PE + 1i	34.67 ± 1.52	41.30 ± 0.84	43.08 ± 0.73	50.78 ± 0.1
PE + 2i	34.67 ± 1.52	41.10 ± 0.67	42.46 ± 0.57	46.67 ± 0.52
PE + Ag + C	34.67 ± 1.52	38.49 ± 1.23	38.83 ± 1.09	41.28 ± 0.49
PE + Ag + 1i	34.67 ± 1.52	36.03 ± 1.50	36.15 ± 1.12	37.79 ± 0.63
PE + Ag + 2i	34.67 ± 1.52	34.89 ± 0.83	35.46 ± 0.37	36.72 ± 0.08
Overall	34.67 ± 1.52A	38.07 ± 0.79A	39.79 ± 0.91B	51.37 ± 0.14B

Data are means ± SD of three replicates.

A, B means in the same column with different letters are significantly different ($p \leq .05$; mean separation was performed by Tukey test).

TABLE 4 Effect of different treatments on *B1* value during storage

Treatment	Storage time (Week)			
	0 Week	1 Week	2 Week	3 Week
C	14.62 ± 0.25	17.62 ± 0.31	20.12 ± 0.17	19.54 ± 0.47
PE + C	14.62 ± 0.25	16.62 ± 0.52	18.19 ± 0.22	18.79 ± 0.43
PE + 1i	14.62 ± 0.25	16.18 ± 0.43	17.49 ± 0.91	18.30 ± 0.08
PE + 2i	14.62 ± 0.25	15.31 ± 0.09	16.17 ± 1.03	17.42 ± 0.43
PE + Ag + C	14.62 ± 0.25	15.57 ± 0.12	16.49 ± 0.52	17.73 ± 0.71
PE + Ag + 1i	14.62 ± 0.25	14.82 ± 0.34	15.73 ± 0.72	16.98 ± 0.51
PE + Ag + 2i	14.62 ± 0.25	14.71 ± 0.67	15.40 ± 0.63	16.97 ± 0.49
Overall	14.62 ± 0.25 A	15.34 ± 0.31B	17.22 ± 0.36B	17.52 ± 0.23C

Data are means ± SD of three replicates.

A–C means in the same row with different letters are significantly different ($p \leq .05$; mean separation was performed by Tukey test).

TABLE 5 Effect of different treatments on weight loss value during storage

Treatment	Storage time (Week)			
	0 Week	1 Week	2 Week	3 Week
C	0.05 ± 0.05	0.21 ± 0.09	0.35 ± 0.12	0.49 ± 0.19
PE + C	0.02 ± 0.04	0.17 ± 0.09	0.27 ± 0.15	0.38 ± 0.18
PE + 1i	0.03 ± 0.08	0.19 ± 0.07	0.25 ± 0.09	0.31 ± 0.20
PE + 2i	0.03 ± 0.05	0.18 ± 0.08	0.23 ± 0.08	0.30 ± 0.21
PE + Ag + C	0.02 ± 0.02	0.16 ± 0.06	0.20 ± 0.13	0.29 ± 0.09
PE + Ag + 1i	0.04 ± 0.03	0.20 ± 0.03	0.29 ± 0.15	0.44 ± 0.17
PE + Ag + 2i	0.04 ± 0.09	0.21 ± 0.01	0.31 ± 0.18	0.42 ± 0.26
Overall	0.03 ± 0.05A	0.18 ± 0.13B	0.23 ± 0.5B	0.37 ± 0.07C

Data are means ± SD of three replicates.

A–C means in the same row with different letters are significantly different ($p \leq .05$; mean separation was performed by Tukey test).

volume of the texture (Jaworska & Bernaś, 2010) was more for the irradiated samples in Ag nanoparticles polyethylene films. 276

Elasticity is recovery after removal of the force of the matter that was more in the samples irradiated in Ag nanoparticles polyethylene 278

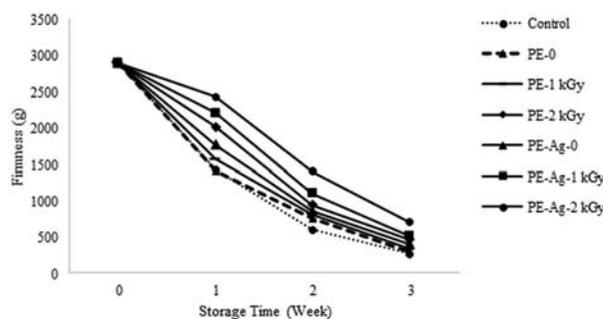


FIGURE 2 Effect of different treatments on firmness value during storage

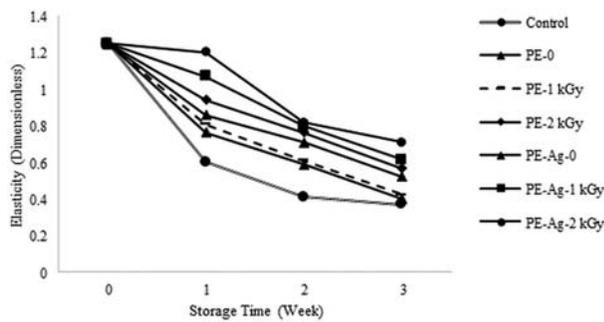


FIGURE 3 Effect of different treatments on elasticity value during storage

TABLE 6 Results of observed colonies on films

Films	Logarithm the number of <i>Escherichia coli</i> × 10 ⁸	Logarithm the number of <i>Staphylococcus aureus</i> × 10 ⁷
PE	2.3 ± 0.07	3.1 ± 0.34
PE + Ag	0.7 ± 0.12	1.2 ± 0.52

280 films during storage. The amount of elasticity is associated with the
 281 elasticity of food (Aday & Caner, 2010). The amount of elasticity in the
 282 control and irradiated sample has reduced during storage time
 F3 283 (Figure 3). The control and nonirradiated sample in polyethylene film
 284 without Ag nanoparticles has less elasticity than the other sample.
 285 Samples of PE-Ag-2 kGy, PE-Ag-1 kGy, PE-2 kGy, and PE-Ag samples
 286 have highest elasticity at the end of study as 0.71 ± 0.0138 , $0.615 \pm$
 287 0.0197 , 0.568 ± 0.037 , and 0.523 ± 0.029 , respectively. The difference
 288 is related to food moisture, minerals and cell water causing turgor pres-
 289 sure that cell water is also influenced by irradiation dose and nanopar-
 290 ticles (Jaworska & Bernaś, 2010).

291 Based on the initial number of bacteria, microbial load in Ag nano-
 T6 292 particles polyethylene films is given in Table 6.

293 Ag nanoparticles reduce the number of colonies of bacteria *S. aureus*
 294 compared with polyethylene film without Ag nanoparticles, but the
 295 bacteria *E. coli* are stronger bacteria (Table 6). Li, Xing, Jiang, Ding, and
 296 Li (2009) showed that Ag and ZnO nanoparticles have antibacterial
 297 properties and the bacteria *E. coli* compared to the bacteria *S. aureus*
 298 are stronger against antibacterial properties that this is due to the dif-
 299 ferences in the bacterial negative-gram and positive-gram structure
 300 and/or dependent on the bacteria sensitivity to hydrogen peroxide
 301 generated from the surface of Ag and ZnO nanoparticles. Conversely,
 302 Ag nanoparticles are an effective way to reduce the microbial load of
 303 negative-gram bacteria such as *E. coli*. The latter results of this study are
 304 consistent with the report of Emamifar et al. (2010).

305 4 | CONCLUSION

306 In this study, combined method of gamma irradiation and Ag nanopar-
 307 ticles polyethylene films, in order to maintain the quality of fresh bot-
 308 tom mushroom, was examined. The results of this study show that the
 309 sample irradiated with a dose of 2 kGy in Ag nanoparticles

polyethylene films makes more proper conditions to maintain the 310
 mushroom. The experimental results and measured parameters such as 311
 pH, weight, color, and textural indices have better values with gamma 312
 irradiation and Ag nanoparticles polyethylene films in these indices. 313
 Also, Ag nanoparticles polyethylene films have antibacterial properties 314
 and compared with conventional films, reduce the accumulation of 315
 microbes and microorganisms. Finally, the results of this study show 316
 that the physical and chemical properties of food irradiated in Ag nano- 317
 particles polyethylene films are maintained and the fresh bottom mush- 318
 room quality are satisfactorily maintained during storage. 319

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ORCID 323

Mahdi Ghasemi-Varnamkhasti  <http://orcid.org/0000-0001-6339-2062> 324
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REFERENCES 326

Abbasi, S., & Azari, S. (2009). Novel microwave-freeze drying of onion 327
 slices. *International Journal of Food Science & Technology*, 44(5), 328
 974–979. 329

Abbott, J. A., & Harker, F. R. (2004). Texture. In K. C. Gross, C. Y. Wang, 330
 & M. Saltweit (Eds.), *The commercial storage of fruits, vegetables, and 331*
florist and nursey stocks. USA: USDA Handbook. 332

Aday, M. S. (2016). Application of electrolyzed water for improving post- 333
 harvest quality of mushroom. *LWT-Food Science and Technology*, 68, 334
 44–51. 335

Aday, M. S., Buyukcan, M. B., & Caner, C. (2013). Maintaining the quality 336
 of strawberries by combined effect of aqueous chlorine dioxide with 337
 modified atmosphere packaging. *Journal of Food Processing and Pres- 338*
ervation, 37(5), 568–581. 339

Aday, M. S., & Caner, C. (2010). Understanding the effects of various 340
 edible coatings on the storability of fresh cherry. *Packaging Technol- 341*
ogy and Science, 23(8), 441–456. 342

Aday, M. S., & Caner, C. (2011). The applications of ‘active packaging 343
 and chlorine dioxide’ for extended shelf life of fresh strawberries. 344
Packaging Technology and Science, 24(3), 123–136. 345

Aday, M. S., Caner, C., & Rahvali, F. (2011). Effect of oxygen and carbon 346
 dioxide absorbers on strawberry quality. *Postharvest Biology and Tech- 347*
nology, 62(2), 179–187. 348

Anthon, G. E., & Barrett, D. M. (2003). Modified method for the determi- 349
 nation of pyruvic acid with dinitrophenylhydrazine in the assessment 350
 of onion pungency. *Journal of the Science of Food and Agriculture*, 83, 351
 1210–1213. 352

Burton, W. G. (1989). *The potato* (3rd ed.). Harlow, UK: Longman Scien- 353
 tific and Technical. 354

Caner, C., & Aday, M. S. (2009). Maintaining quality of fresh strawberries 355
 through various modified atmosphere packaging. *Packaging Technol- 356*
ogy and Science, 22(2), 115–122. 357

Cao, S., Hu, Z., Pang, B., Wang, H., Xie, H., & Wu, F. (2010). Effect of 358
 ultrasound treatment on fruit decay and quality maintenance in 359
 strawberry after harvest. *Food Control*, 21(4), 529–532. 360

Charlesby, A. (2016). *Atomic radiation and polymers: International series of 361*
monographs on radiation effects in materials. Elsevier. 362

AQ1

AQ2

- 363 Choi, D. S., Park, S. H., Choi, S. R., Kim, J. S., & Chun, H. H. (2015). The
364 combined effects of ultraviolet-C irradiation and modified atmos-
365 phere packaging for inactivating *Salmonella enterica* serovar Typhimur-
366 ium and extending the shelf life of cherry tomatoes during cold
367 storage. *Food Packaging and Shelf Life*, 3, 19–30.
- 368 Donglu, F., Wenjian, Y., Kimatu, B. M., Mariga, A. M., Liyan, Z., Xinxin, A.,
369 & Qiuhui, H. (2016). Effect of nanocomposite-based packaging on
370 storage stability of mushrooms (*Flammulina velutipes*). *Innovative Food
371 Science & Emerging Technologies*, 33, 489–497.
- 372 Echegoyen, Y., & Nerin, C. (2015). Performance of an active paper based
373 on cinnamon essential oil in mushrooms quality. *Food Chemistry*, 170,
374 30–36.
- 375 Ekezie, F. G. C., Cheng, J. H., & Sun, D. W. (2018). *Effects of nonthermal
376 food processing technologies on food allergens: A review of recent
377 research advances*. Trends in Food Science & Technology.
- 378 Emamifar, A., Kadivar, M., Shahedi, M., & Soleimani-Zad, S. (2010).
379 Evaluation of nanocomposite packaging containing Ag and ZnO on
380 shelf life of fresh orange juice. *Innovative Food Science & Emerging
381 Technologies*, 11(4), 742–748.
- 382 Fernandes, Á., Barreira, J. C., Antonio, A. L., Oliveira, M. B. P., Martins,
383 A., & Ferreira, I. C. (2016). Extended use of gamma irradiation in
384 wild mushrooms conservation: Validation of 2 kGy dose to pre-
385 serve their chemical characteristics. *LWT-Food Science and Technol-
386 ogy*, 67, 99–105.
- 387 Gantner, M., Guzek, D., Pogorzelska, E., Brodowska, M., Wojtasik-Kali-
388 nowska, I., & Godziszewska, J. (in press). The effect of film type and
389 modified atmosphere packaging with different initial GAS composi-
390 tion on the shelf life of white mushrooms (*Agaricus bisporus* L.). *Jour-
AQ3 391 nal of Food Processing and Preservation*.
- 392 Gilman, J., Jacxsens, L., Meulenaer, D., B., & Devlieghere, F. (2015).
393 Modified atmosphere packaging and irradiation to preserve contem-
394 porary food-based art: An experimental study. *Journal of Cultural Her-
395 itage*, 16(3), 391–397.
- 396 Guan, W., Fan, X., & Yan, R. (2013). Effect of combination of ultraviolet
397 light and hydrogen peroxide on inactivation of *Escherichia coli* O157:
398 H7, native microbial loads, and quality of button mushrooms. *Food
399 Control*, 34(2), 554–559.
- 400 Han, L., Qin, Y., Liu, D., Chen, H., Li, H., & Yuan, M. (2015). Evaluation of
401 biodegradable film packaging to improve the shelf-life of *Boletus
402 edulis* wild edible mushrooms. *Innovative Food Science & Emerging
403 Technologies*, 29, 288–294.
- 404 Jaworska, G., & Bernaś, E. (2010). Effects of pre-treatment, freezing and
405 frozen storage on the texture of *Boletus edulis* (Bull: Fr.) mushrooms.
406 *International Journal of Refrigeration*, 33(4), 877–885.
- 407 Jo, C., Son, J. H., Shin, M. G., & Byun, M. W. (2003). Irradiation effects
408 on color and functional properties of persimmon (*Diospyros
409 kaki* L. folium) leaf extract and licorice (*Glycyrrhiza Uralensis*
410 Fischer) root extract during storage. *Radiation Physics and Chemis-
411 try*, 67(2), 143–148.
- 412 Khan, F. M., & Gibbons, J. P. (2014). *Khan's the physics of radiation ther-
413 apy*. Lippincott Williams & Wilkins.
- 414 Khatun, A., Hossain, A., Islam, M., Munshi, K., Akter, A., Rahman, B., &
415 Huque, R. (2017). Evaluation of gamma irradiation and boiling treat-
416 ment on antioxidant status in different spices. *Journal of Food Process
417 Engineering*, 40(3), e12482.
- 418 Kim, K. M., Ko, J. A., Lee, J. S., Park, H. J., & Hanna, M. A. (2006).
419 Effect of modified atmosphere packaging on the shelf-life of
420 coated, whole and sliced mushrooms. *LWT-Food Science and Tech-
421 nology*, 39(4), 365–372.
- 422 Koutsimanis, G., Harte, J., & Almenar, E. (2015). Development and evalu-
423 ation of a new packaging system for fresh produce: A case study on
fresh cherries under global supply chain conditions. *Food and Biopro-
cess Technology*, 8(3), 655–669. 424 425
- Li, X., Xing, Y., Jiang, Y., Ding, Y., & Li, W. (2009). Antimicrobial activities
of ZnO powder-coated PVC film to inactivate food pathogens. *Inter-
national Journal of Food Science & Technology*, 44(11), 2161–2168. 426 427 428
- Madera-Santana, T. J., Meléndrez, R., González-García, G., Quintana-
Owen, P., & Pillai, S. D. (2016). Effect of gamma irradiation on physico-
chemical properties of commercial poly (lactic acid) clamshell for
food packaging. *Radiation Physics and Chemistry*, 123, 6–13. 429 430 431 432
- Mahrouz, M., Lacroix, M., D'aprano, G., Oufedjikh, H., Boubekri, C., &
Gagnon, M. (2002). Effect of γ -irradiation combined with washing
and waxing treatment on physicochemical properties, vitamin C, and
organoleptic quality of *Citrus clementina* Hort. ex. Tanaka. *Journal of
Agricultural and Food Chemistry*, 50(25), 7271–7276. 433 434 435 436 437
- Majeed, A., Muhammad, Z., Ullah, R., Ullah, Z., Ullah, R., Chaudhry, Z., &
Siyar, S. (2017). Effect of gamma irradiation on growth and post-
harvest storage of vegetables. *PSM Biological Research*, 2(1), 30–35. 438 439 440
- Mami, Y., Peyvast, G., Ziaie, F., Ghasemnezhad, M., & Salmanpour, V.
(2014). Improvement of shelf life and postharvest quality of white
button mushroom by electron beam irradiation. *Journal of Food Proc-
essing and Preservation*, 38(4), 1673–1681. 441 442 443 444
- Marra, A., Boumail, A., Cimmino, S., Criado, P., Silvestre, C., & Lacroix, M.
(2016). Effect of PLA/ZnO packaging and gamma radiation on the content
of *Listeria innocua*, *Escherichia coli* and *Salmonella enterica* on Ham during
Storage at 4 °C. *Journal of Food Science and Engineering*, 6, 245–259. 445 446 447 448
- Oliveira, F., Sousa-Gallagher, M. J., Mahajan, P. V., & Teixeira, J. A.
(2012a). Development of shelf-life kinetic model for modified atmos-
phere packaging of fresh sliced mushrooms. *Journal of Food Engineer-
ing*, 111(2), 466–473. 449 450 451 452
- Oliveira, F., Sousa-Gallagher, M. J., Mahajan, P. V., & Teixeira, J. A.
(2012b). Evaluation of MAP engineering design parameters on qual-
ity of fresh-sliced mushrooms. *Journal of Food Engineering*, 108(4),
507–514. 453 454 455 456
- Oufedjikh, H., Mahrouz, M., Amiot, M. J., & Lacroix, M. (2000). Effect of
 γ -irradiation on phenolic compounds and phenylalanine ammonia-
lyase activity during storage in relation to peel injury from peel of
Citrus clementina Hort. Ex. Tanaka. *Journal of Agricultural and Food
Chemistry*, 48(2), 559–565. 457 458 459 460 461
- Oufedjikh, H., Mahrouz, M., Lacroix, M., Amiot, M. J., & Taccini, M.
(1998). The influence of gamma irradiation on flavonoids content
during storage of irradiated clementina. *Radiation Physics and Chemis-
try*, 52(1–6), 107–112. 462 463 464 465
- Qin, Y., Liu, D., Wu, Y., Yuan, M., Li, L., & Yang, J. (2015). Effect of PLA/
PCL/cinnamaldehyde antimicrobial packaging on physicochemical and
microbial quality of button mushroom (*Agaricus bisporus*). *Postharvest
Biology and Technology*, 99, 73–79. 466 467 468 469
- Restrepo-Flórez, J. M., Bassi, A., & Thompson, M. R. (2014). Microbial
degradation and deterioration of polyethylene—A review. *Internation-
al Biodeterioration & Biodegradation*, 88, 83–90. 470 471 472
- Sapers, G. M., Miller, R. L., Choi, S. W., & Cooke, P. H. (1999). Structure
and composition of mushrooms as affected by hydrogen peroxide
wash. *Journal of Food Science*, 64(5), 889–892. 473 474 475
- Schmid, M., Held, J., Hammann, F., Schlemmer, D., & Noller, K. (2015).
Effect of UV/radiation on the packaging-related properties of whey
protein isolate based films and coatings. *Packaging Technology and
Science*, 28(10), 883–899. 476 477 478 479
- Severino, R., Ferrari, G., Vu, K. D., Donsi, F., Salmieri, S., & Lacroix, M.
(2015). Antimicrobial effects of modified chitosan based coating con-
taining nanoemulsion of essential oils, modified atmosphere packag-
ing and gamma irradiation against *Escherichia coli* O157: H7 and
Salmonella typhimurium on green beans. *Food Control*, 50, 215–222. 480 481 482 483 484

485 Taghizadeh, M., Gowen, A., Ward, P., & O'donnell, C. P. (2010). Use of
486 hyperspectral imaging for evaluation of the shelf-life of fresh white but-
487 ton mushrooms (*Agaricus bisporus*) stored in different packaging films.
488 *Innovative Food Science & Emerging Technologies*, 11(3), 423–431.

489 Wong, K. M., Decker, E. A., Autio, W. R., Toong, K., DiStefano, G., & Kin-
490 chla, A. J. (2017). Utilizing mushrooms to reduce overall sodium in
491 taco filling using physical and sensory evaluation. *Journal of Food Sci-
492 ence*, 82(10), 2379–2386.

493 Xu, N., Hu, X., Xu, W., Li, X., Zhou, L., Zhu, S., & Zhu, J. (2017). Mush-
494 rooms as efficient solar steam-generation devices. *Advanced Materi-
495 als*, 29(28), 1606762.

496 Yuk, H. G., Yoo, M. Y., Yoon, J. W., Marshall, D. L., & Oh, D. H. (2007).
497 Effect of combined ozone and organic acid treatment for control of
498 *Escherichia coli* O157: H7 and *Listeria monocytogenes* on enoki mush-
499 room. *Food Control*, 18(5), 548–553.

500 Yurttas, Z. S., Moreira, R. G., & Castell-Perez, E. (2014). Combined vac-
501 uum impregnation and electron-beam irradiation treatment to extend
the storage life of sliced white button mushrooms (*Agaricus bisporus*).
Journal of Food Science, 79(1), 39–46.

Zhou, L., Lv, S., He, G., He, Q., & Shi, B. I. (2011). Effect of PE/AG2O
nanopackaging on the quality of apple slices. *Journal of Food Quality*,
34(3), 171–176.

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